# Evaluation of two engineered antibody fragments, derived from the human anti-CD20 Ofatumumab, as tracers for immunoPET

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### Abstract

**Background:** CD20 is a cell surface antigen that is expressed on the majority of B-cell lymphomas and B-cell leukemias. Several monoclonal antibodies (mAbs), including the fully human IgG1, Ofatumumab, have been used successfully to target human CD20. Ofatumumab is FDA-approved for the treatment of patients with chronic lymphocytic leukemia (CLL) and is under ongoing clinical development for other hematological malignancies and autoimmune diseases. In this study Ofatumumab was engineered into smaller fragments including a minibody (scFv-C<sub>H</sub>3; 80kDa) and a cys-diabody (scFv dimer; 50kDa) to enhance pharmacokinetic parameters and optimize possible application for immunoPET imaging.

Methods: The variable (V) genes of Ofatumumab were reformatted into two minibodies (Mb) and four cys-diabodies (Cys-Db), differing in orientation of the V genes and linker lengths. Following in vitro characterization of each variant, a lead candidate from each format was produced at larger scale for *in vivo* evaluation. Ofatumumab, Mb and Cys-Db were radioiodinated with I-131 (t<sub>1/2</sub> 8d) for biodistribution and I-124 (t<sub>1/2</sub> 4.2d) for PET imaging studies using the lodogen method. Female SCID mice bearing Ramos (CD20<sup>+</sup>) and BC-1 (CD20<sup>-</sup>) tumors were used. Blocking studies were performed by pre-injecting 0.2-1mg of unlabeled Ofatumumab 24h prior to imaging. The percent injected dose per gram (%ID/g) was determined in harvested tumor, blood and organs.

**Results:** The lead Cys-Db and Mb were shown to have similar binding affinity (0.8 nM) to that of the parental Ofatumumab. Receptor specific binding was confirmed by competitive cell binding assays using both fluorophore-conjugated and radiolabeled antibody fragments. Blood clearances resulted in MRTs of 3.8h, 8.3h and 24.9h for Cys-Db, Mb and Ofatumumab, respectively. Due to its slower initial distribution, the Mb had a higher blood AUC (306%ID/g\*h) than that of Ofatumumab (234%) ID/g\*h) and Cys-Db (102% ID/g\*h). Biodistribution studies of Ofatumumab revealed significant uptake in spleen, bone and liver, which was attributed to Fc-mediated clearance as this was not observed with the antibody fragments. Imaging studies with Mb and Cys-Db showed clear delineation of Ramos xenografts; 1.3% ID/g Cys-Db at 8h and 2.7% ID/g Mb at 24h. This resulted in a tumor to blood ratio of 1.7 for the Cys-Db at 8h and 1.1 for the Mb at 24h. Prior administration of excess unlabeled Ofatumumab reduced tumor uptake of the Cys-Db by 50% at 8h and decreased uptake of the Mb by 17% at 24h.

**Conclusion:** Our preliminary results show that antibody fragments derived from Ofatumumab can be used to target CD20 positive cells in vitro and in vivo. The favorable pharmacokinetics of the Cys-Db resulted in higher tumor to blood ratios at 8h and 24h compared to the Mb or Ofatumumab and support the further development of this fragment as a CD20-targeted immunoPET agent.

### **Background and Rationale**

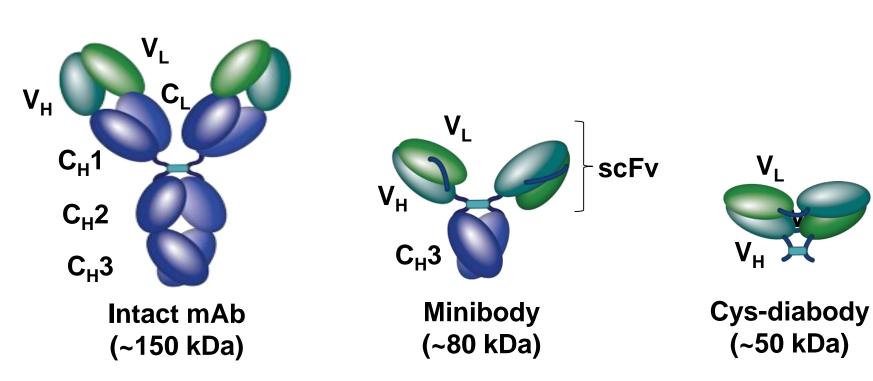
• Ofatumumab, a human monoclonal antibody directed to CD20, was approved in 2009 for the treatment of refractory chronic lymphocytic leukemia.

• CD20 is a B-cell–specific differentiation antigen that is expressed on mature B cells and in most B-cell non-Hodgkin's lymphomas but not on early B-cell progenitors or later mature plasma cells.

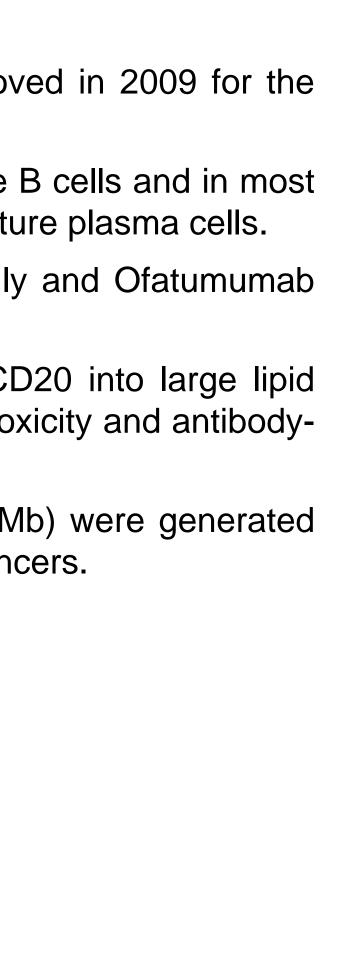
• The CD20 antigen is a member of the membrane-spanning 4A gene family and Ofatumumab binds both the large and small extracellular loops.

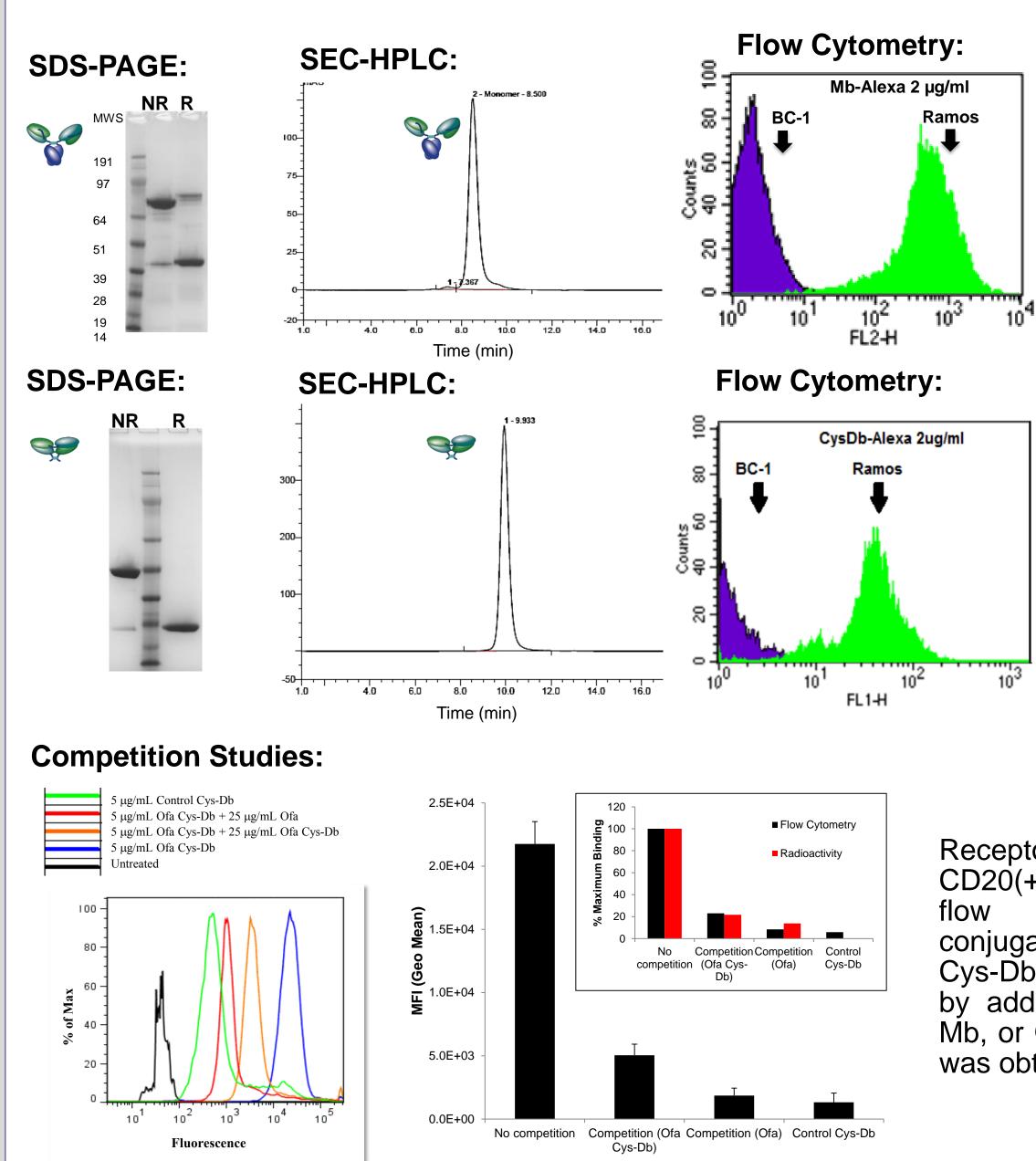
• Type I antibodies (Rituximab and Ofatumumab) induce redistribution of CD20 into large lipid rafts in the plasma membrane and confer strong complement-dependent cytotoxicity and antibodydependent cell-mediated cytotoxicity but have minimal direct antitumor effects.

• In this study a diabody with terminal cysteines (Cys-Db) and a Minibody (Mb) were generated from of atumumab and evaluated for their suitability to image CD20 positive cancers.



Schematics of an intact monoclonal antibody (mAb), a minibody and a Cys-Db. The various domains are indicated. Molecular weights are shown below. V = variable, C = constant, L = light, H = heavy

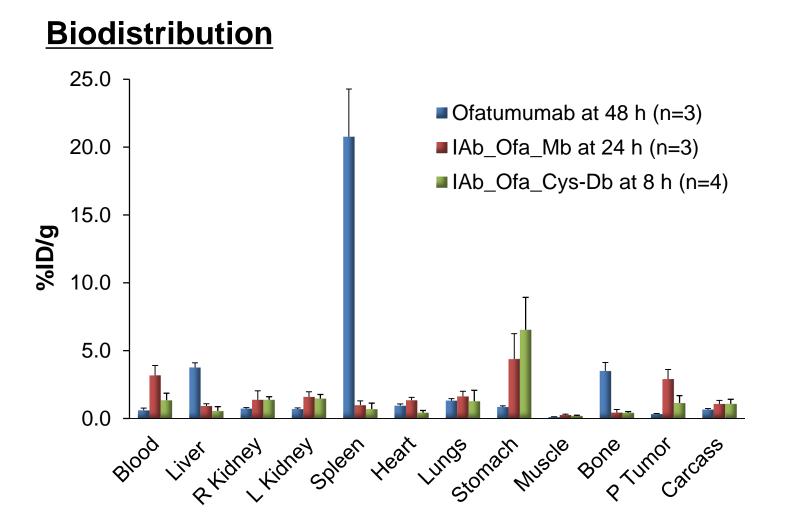




# In Vivo Biodistribution and Pharmacokinetics

The fragments and Ofatumumb were radioidinated with I-131 using the lodogen method. High immunoreactivity to Ramos cell were maintained after the labeling

Radiochemical Purity Specific Activity (µCi/µg) Immunoreactivity



Groups of 3-4 mice were injected i.v. with 10-14 µCi of I-131-labeled Cys-Db, Mb or Ofatumumb. Biodistributions were performed at 8, 25 and 48 hrs for the Cys-Db, Mb and Ofatumumab, respectively. At specific time-points tails were nicked with a scalpel and 10 µL blood collected and counted in a gamma counter. The activity was decay corrected and the %ID/g was calculated. The MRTs from the blood clearance profiles shown above, were determined to be 2.9h, 6.2h and 13.2h for the Cys-Db, Mb and Ofatumumab, respectively. Due to its' slower initial distribution, the Mb had a higher blood AUC (277%ID/g\*h) than that of Ofatumumab (212% ID/g\*h) and Cys-Db (100% ID/g\*h) in this experiment. The iodistribution shows that Ofatumumab has high uptake in spleen, liver and bone which is probably due to Fc-mediated clearance.

## In Vitro Characterization

Supernatants from large scale production of the Mb and Cys-Db in mammalian cell culture were purified by ion exchange chromatography followed by ceramic hydroxyapatite. Purity and aggregation of the Mb and Cys-Db were evaluated by SDS-PAGE and size exclusion chromatography (SE-HPLC). Binding was evaluated by flow cytometry. Receptor-specific binding was evaluated by competition studies.

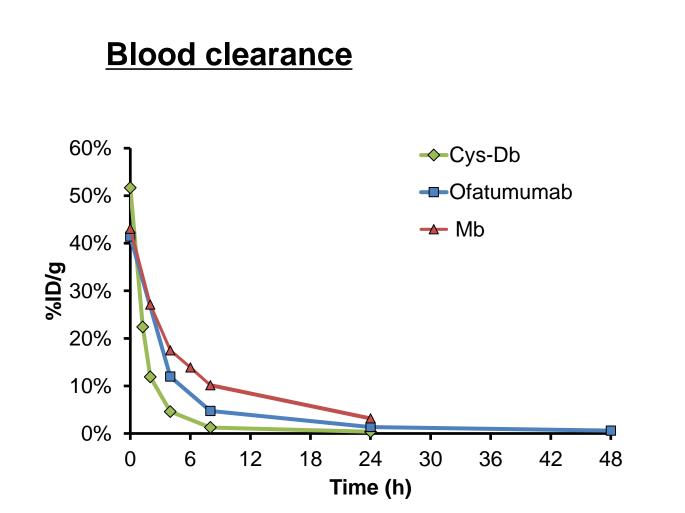
> SDS-PAGE showed that the Mb and Cys-Db migrated as covalent dimers of ~80 kDa and 50 kDa, respectively. In the presence of a reducing agent they monomers of ~40 kDa and 25 kDa. The purity by SDS-PAGE was >90%.

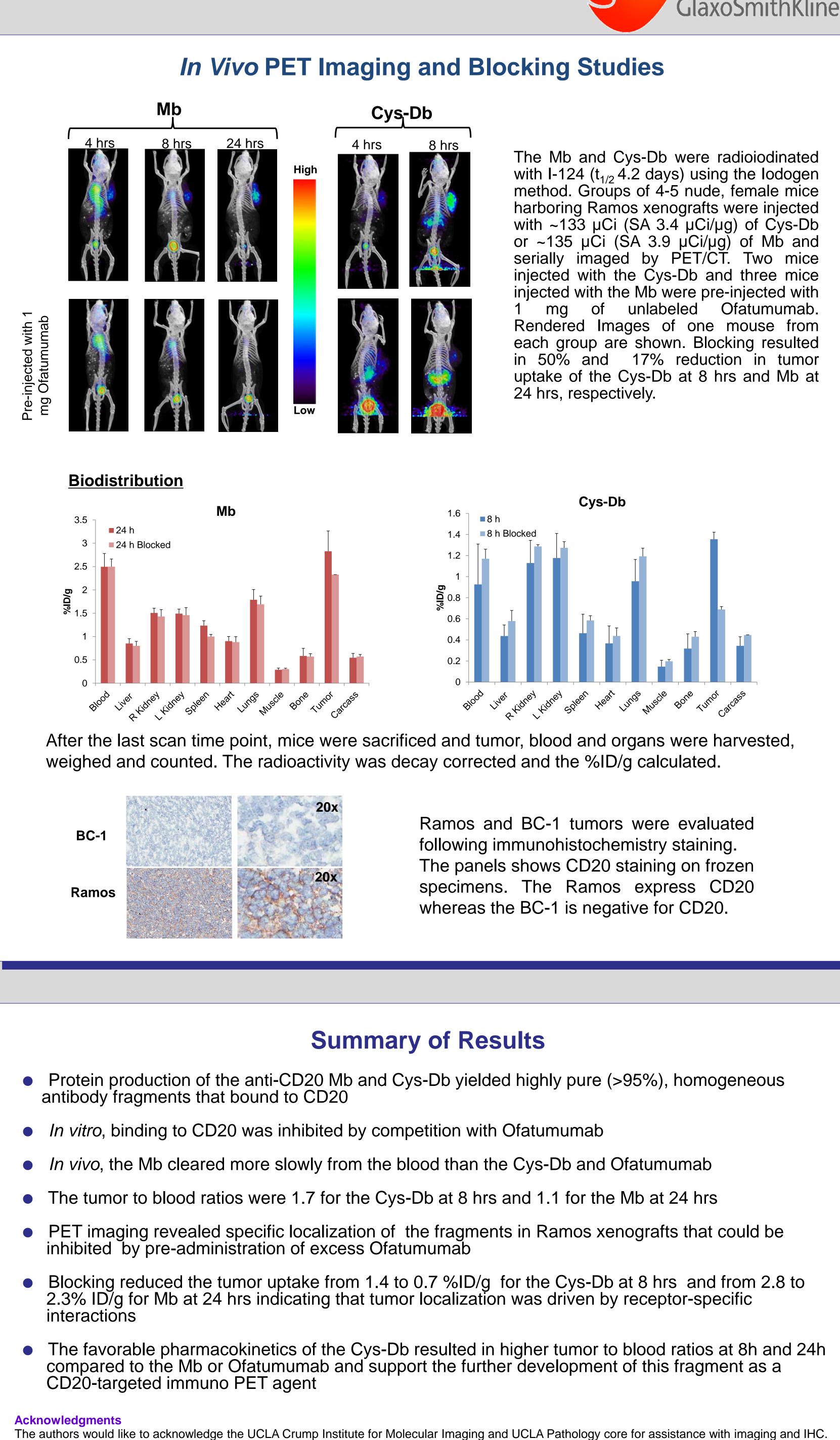
SE-HPLC analysis showed that both proteins eluted as a single peak corresponding to dimers of ~80 kDa and 50 kDa in size. The purity by SEC was estimated to be >98% for both fragments.

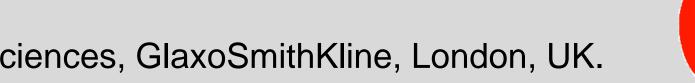
confirmed cytometry binding to CD20 expressing Ramos cells, and not to the negative BC-1 cells.

Receptor-specific binding of Cys-Db to CD20(+ve) Ramos cells was confirmed by after fluorophore conjugation and radioiodination. Binding of Cys-Db to Ramos cells could be inhibited by addition of excess unlabeled Cys-Db, Mb, or Ofatumumab. Similar displacement was obtained with the Mb (not shown)

Cys-Db x2	Mb	Ofatumumab
99.3%	96.8%	96.4%
3.33/3.33	3.57	3.56
66.3 vs.4.0%	72.7 vs. 5.7%	83.5 vs. 2.7%







The Mb and Cys-Db were radioiodinated with I-124 ( $t_{1/2}$  4.2 days) using the lodogen method. Groups of 4-5 nude, female mice harboring Ramos xenografts were injected with ~133 µCi (SA 3.4 µCi/µg) of Cys-Db or ~135  $\mu$ Ci (SA 3.9  $\mu$ Ci/ $\mu$ g) of Mb and serially imaged by PET/CT. Two mice injected with the Cys-Db and three mice injected with the Mb were pre-injected with Ofatumumab. Images of one mouse from each aroup are shown. Blocking resulted 50% and 17% reduction in tumor uptake of the Cys-Db at 8 hrs and Mb at 24 hrs, respectively.

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